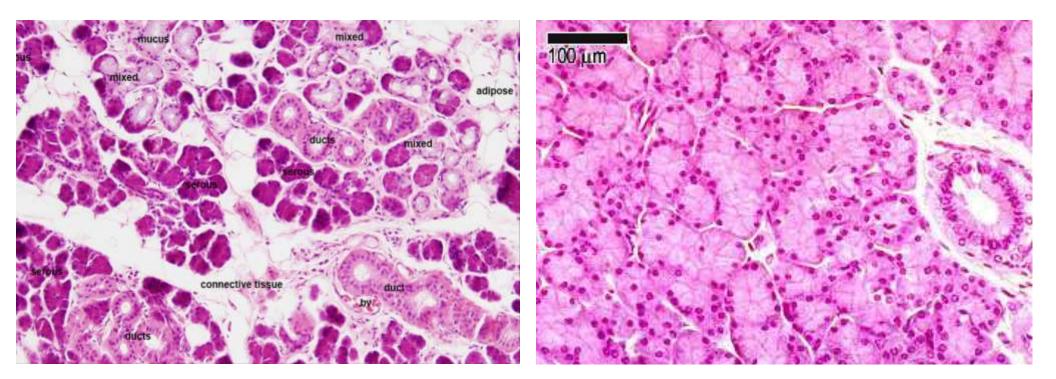
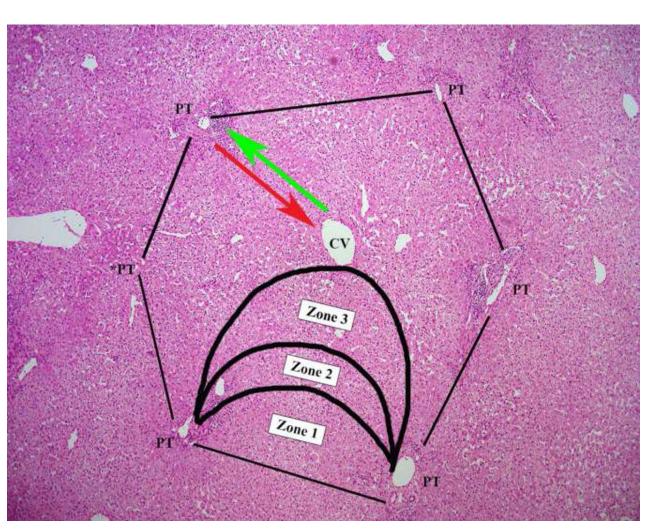
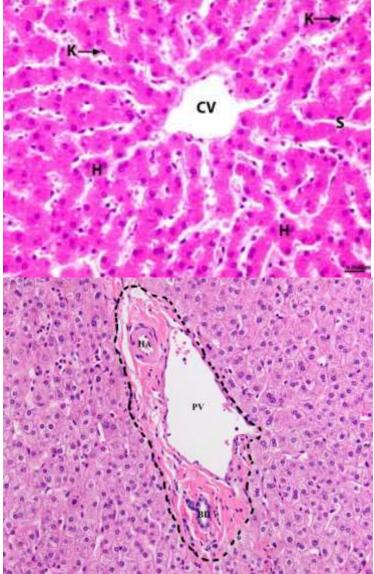
- •Study of histological slides: Salivary gland, Liver, Pancreas, Stomach and Intestine.
- •Glucose estimation
- •Amylase assay in the given sample
- •Determination of blood groups (ABO and Rh factor)
- •Erythrocyte counting
- •Total leucocytes counting in blood
- •Study of histological slides: Kidney, Heart and Lungs
- •Study of Kidney, Heart and Lungs with models/PPT

Study of histological slides: Salivary gland

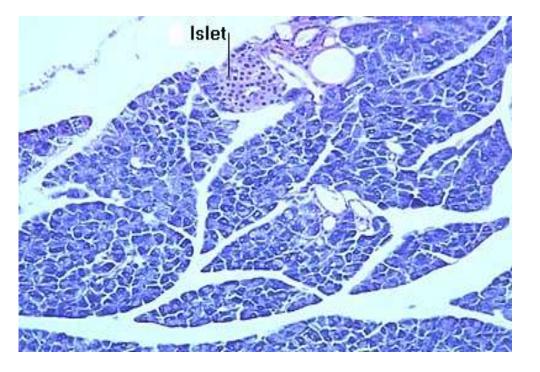


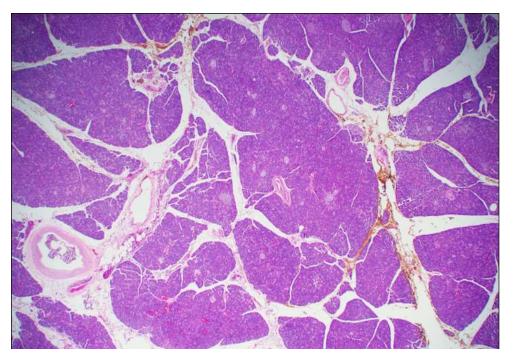
Study of histological slides: Liver



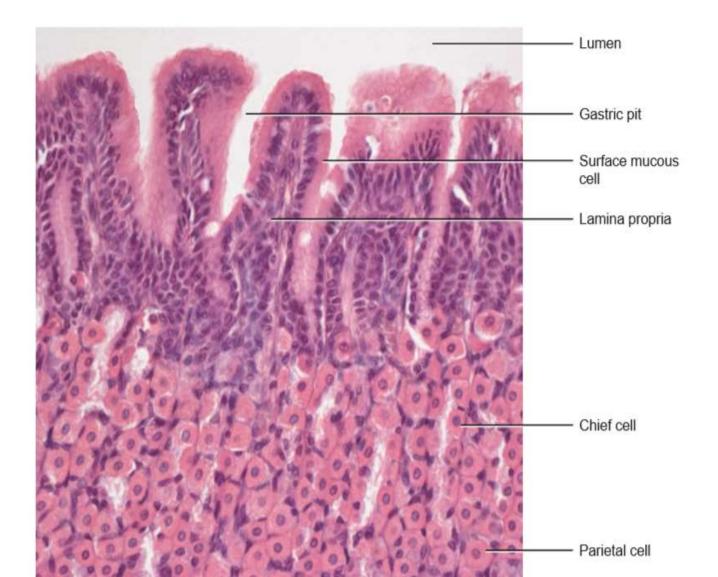


Study of histological slides: Pancreas

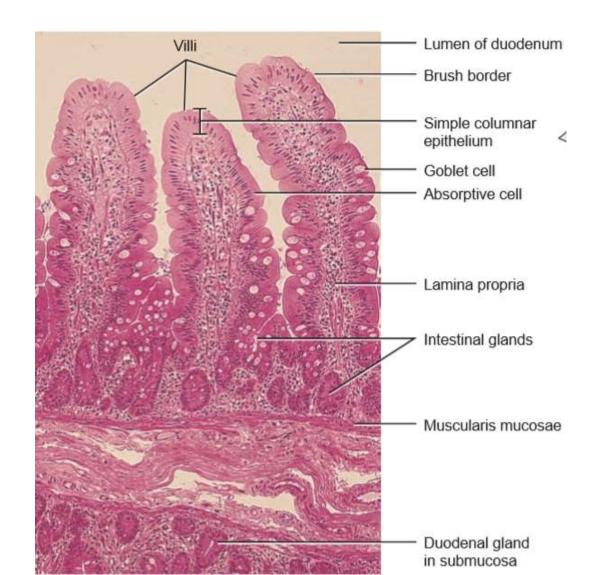




Study of histological slides: Stomach



Study of histological slides: Small Intestine



Determination of blood groups (ABO and Rh factor)

Objective:

To determine an individual's ABO blood group (A, B, AB, or O) and Rh factor (positive or negative).

Materials: Blood samples from the individual to be tested, Anti-A serum, Anti-B serum, Anti-D (Rh) serum, Microscope slides, Disposable lancets or sterile needles, Alcohol swabs, Safety gloves, Droppers

Procedure:

Sterilize the fingertip using an alcohol swab.

Prick the fingertip with a disposable lancet or sterile needle.

Allow a few drops of blood to flow and collect them in separate droppers or directly onto the labeled agglutination cards/test tubes.

Forward Blood Grouping (ABO):

Place one drop of the individual's blood on three separate microscope slides.

Add one drop of Anti-A serum to the blood on the first slide, Anti-B serum to the second slide, and Anti-D (Rh) serum to the third slide.

Determination of blood groups (ABO and Rh factor)

Mix each slide gently. After mixing, tilt each slide back and forth to observe for agglutination. If clumping occurs in the Anti-A serum-added slide, the blood group is A. If clumping occurs in the Anti-B serum-added slide, the blood group is B. If clumping occurs in both Anti-A and Anti-B serum-added slides, the blood group is AB. If no clumping occurs in either Anti-A or Anti-B serum-added slides, the blood group is O.

Rh (D) Factor Testing:

Examine the third slide where Anti-D (Rh) serum was added. If clumping occurs, the Rh factor is positive (+). If no clumping occurs, the Rh factor is negative (-).

Recording Results:

Document the individual's blood group and Rh factor based on the observed agglutination patterns.

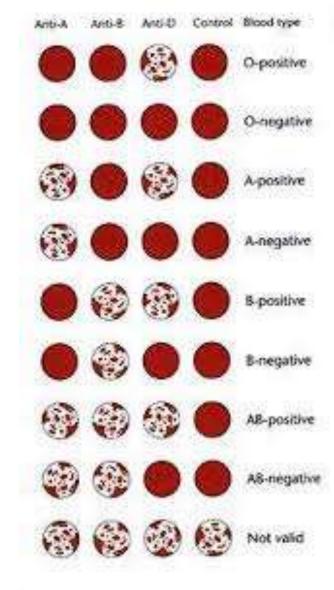
Precautions:

Always wear safety gloves when handling blood samples to prevent contamination and potential exposure to blood-borne pathogens.

Dispose of lancets, needles, and any used materials in a designated biohazard waste container.

Determination of blood groups (ABO and Rh factor)

	Group A	Group B	Group AB	Group O
Red blood cell type			AB	
Antibodies in plasma	、 、 、 、 、 、 、 、 、 、 、 、 、 、	Anti-A	None	Anti-A and Anti-B
Antigens in red blood cell	₽ A antigen	↑ B antigen	P↑ A and B antigens	None



Erythrocyte counting

Objective:

To determine the concentration of red blood cells per unit volume of blood.

Materials:

Microscope, Hemocytometer, Cover slip, Pipette, Diluting fluid (e.g., isotonic saline or Hayem's solution), Lancet or needle, Microscope slides

Procedure:

Preparation of Diluting Fluid:

Prepare a diluting fluid by mixing isotonic saline or Hayem's solution with distilled water as per the manufacturer's instructions or standard laboratory protocol.

Sample Collection:

Sterilize the fingertip using an alcohol swab.

Prick the fingertip with a lancet or needle.

Collect a small drop of blood without squeezing the fingertip excessively.

Preparation of Blood Sample:

Wipe away the first drop of blood.

Fill the pipette with blood up to the 0.5 mark.

Mix blood with 0.5 mL of diluting fluid by gently squeezing and releasing the pipette several times.

Erythrocyte counting

Loading the Neubauer Chamber:

Clean the surface of the Neubauer chamber and cover slip to remove any dust or debris. Place the cover slip over the counting chamber of the Neubauer chamber without trapping air bubbles.

Allow the blood to settle for a few minutes to ensure an even distribution in the counting chamber.

Counting RBCs:

Count the number of red blood cells in multiple grids to get an average count. Ensure that you count only the cells that lie within the grid lines or on the top and right lines of each grid.

Recording Results:

Record the RBC count in millions per microliter ($x10^{6}/\mu L$).

Safety Precautions:

Handle all biological samples with care to avoid contamination and potential exposure to bloodborne pathogens.

Dispose of lancets, needles, and any used materials in a designated biohazard waste container.

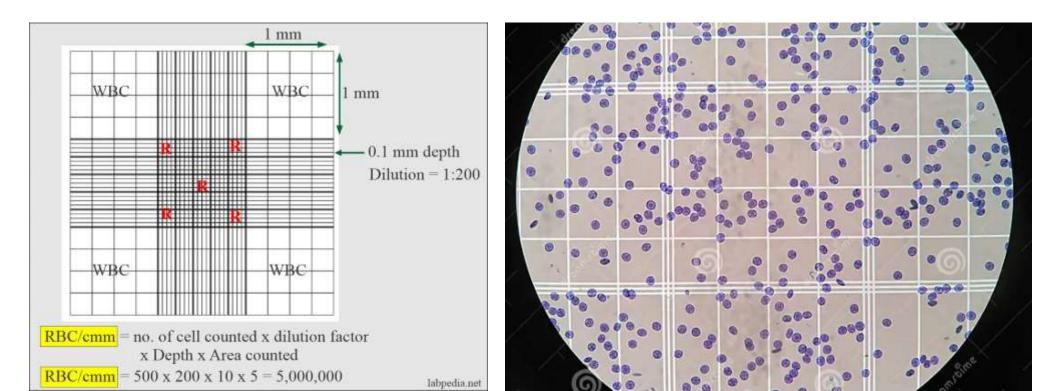
Erythrocyte counting

Calculating RBC Count:

• Use the formula:

 $RBC \ count = \frac{Average \ number \ of \ RBCs \ counted}{Number \ of \ squares \ counted} \times Dilution \ factor$

- Common dilution factors:
 - Unopette method: 200
 - Hayem's method: 10



Total leucocytes counting in blood

Procedure:

Fill the blood into the 0.5 marks and then add the TLC solution.

Fill the pipette with the TLC solution to point 11.

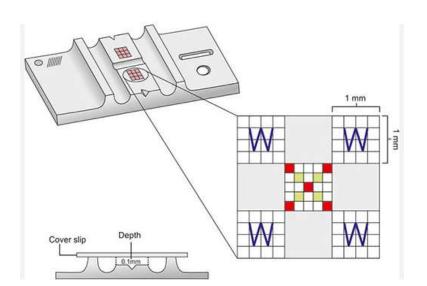
Remove the rubber tubing.

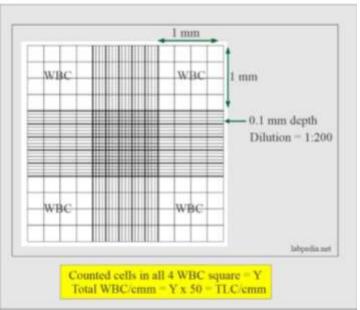
Seal both ends or hold in between two fingers.

Shake for 1 minute or preferably for 2 minutes.

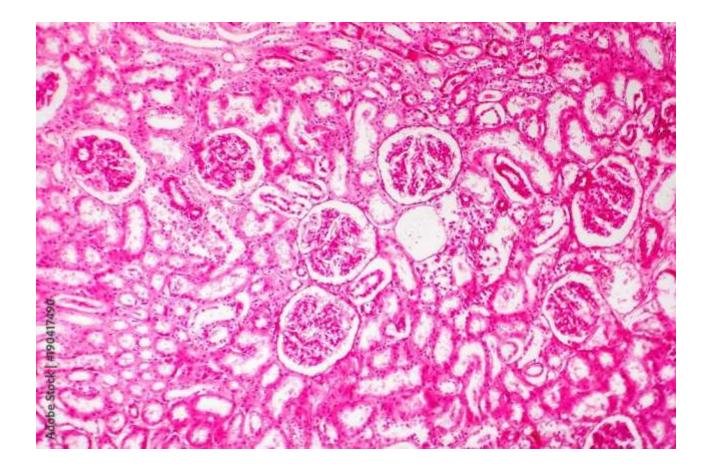
After thorough mixing, discard the first few drops and then gently fill the chamber until the platform is filled.

Allow the chamber on the microscope stage for 2 to 3 minutes till the cells are settled.

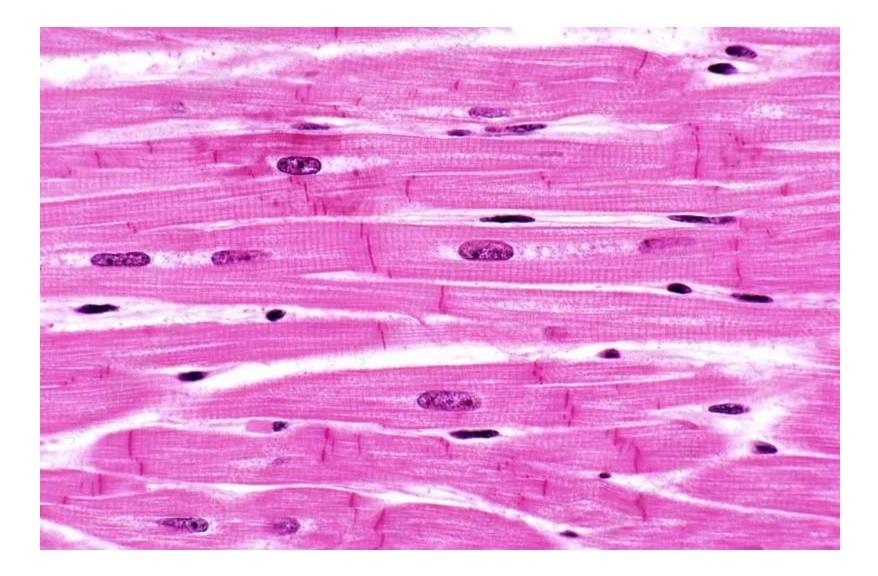




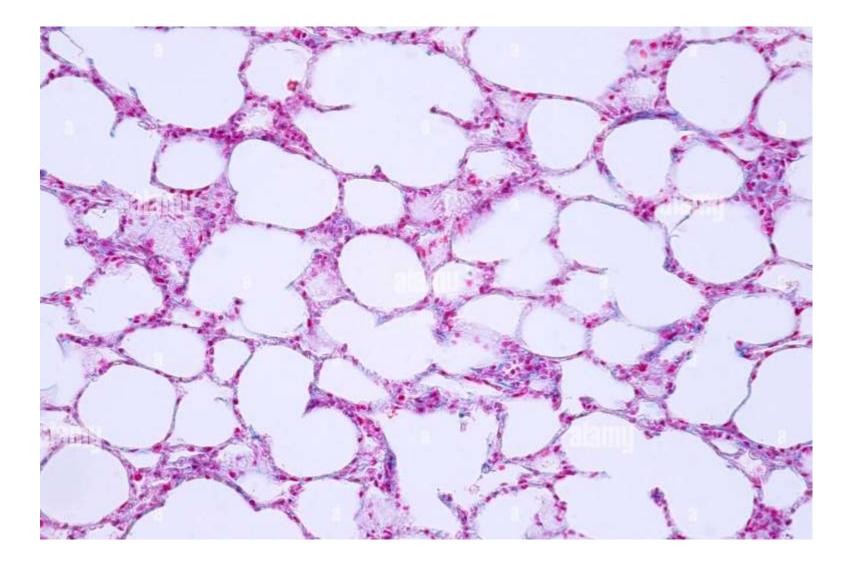
Study of histological slides: Kidney



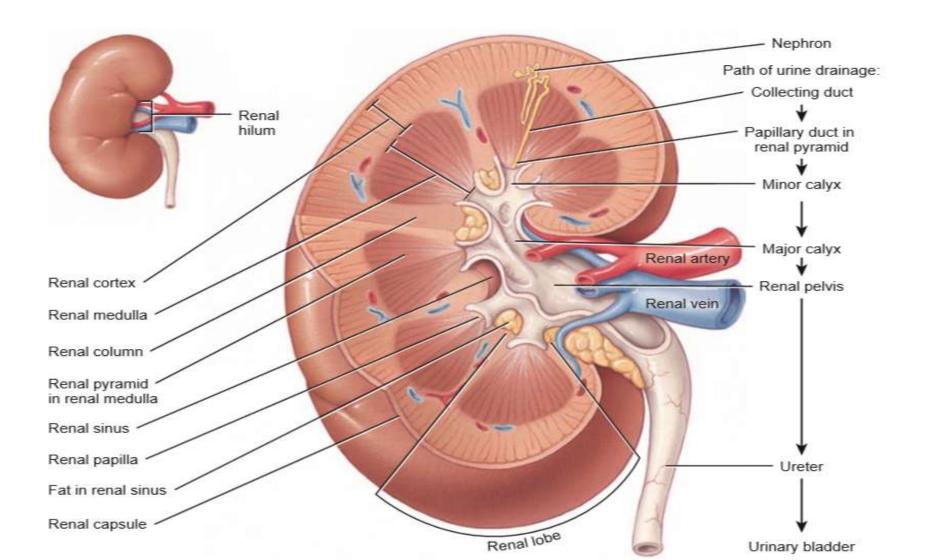
Study of histological slides: Heart



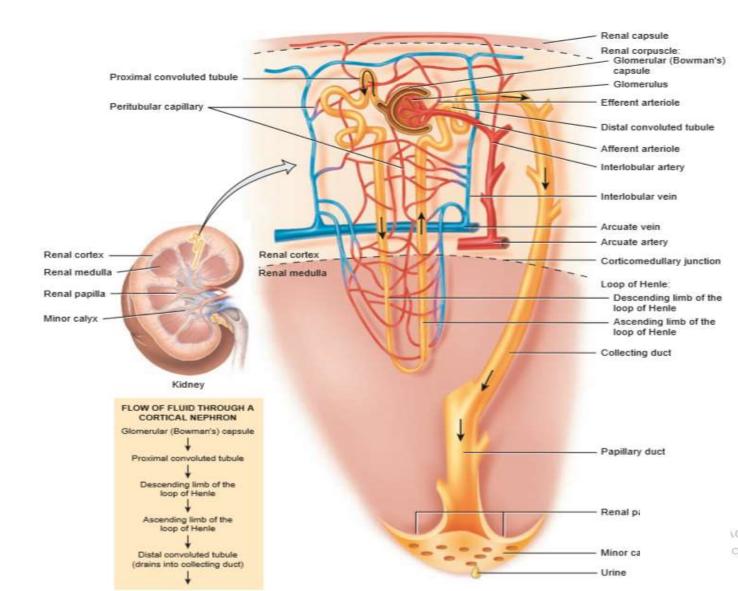
Study of histological slides: Lungs



Study of Kidney with models/PPT

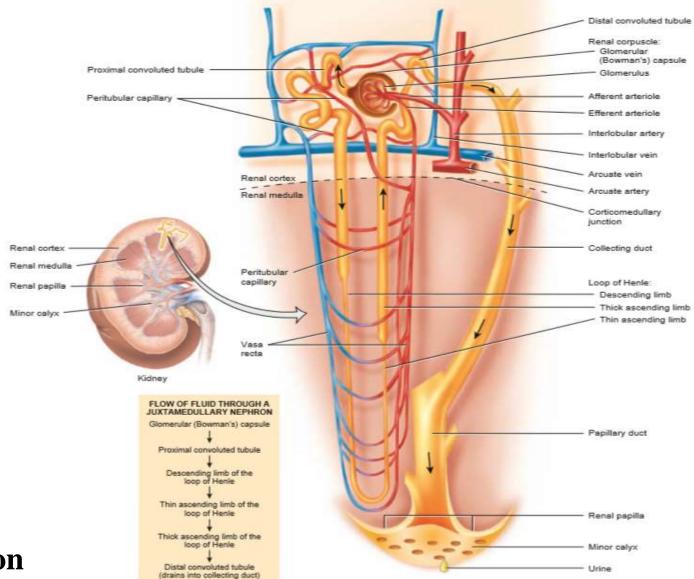


Study of Kidney with models/PPT



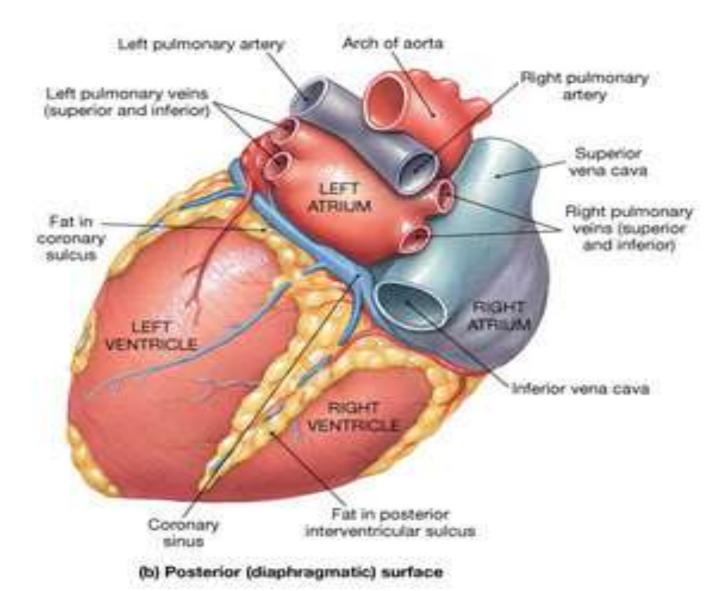
Cortical Nephron

Study of Kidney with models/PPT

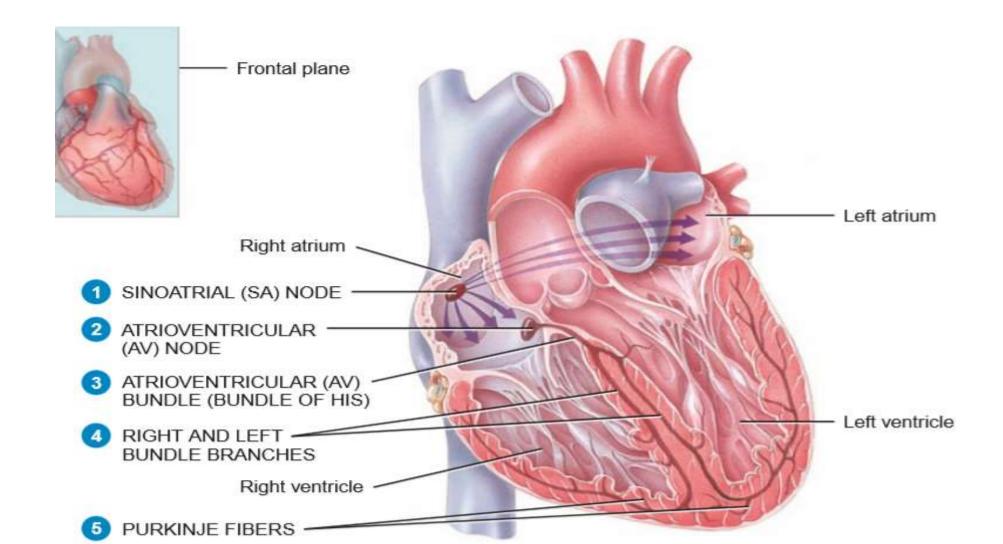


Juxtamedullary Nephron

Study of Heart with models/PPT



Study of Heart with models/PPT



Study of Lungs with models/PPT

